Development of NOG mice



- 1. T and B cell deficient
- 2. NK cell deficient
- 3. Reduced macrophage and dendritic cell function
- 4. Complement activity deficient
- 5. No incidence of lymphoma
- 6. No T, B cell leakiness
- 7. Long life span
- 8. Sensitive for irradiation
- 9. Sensitive against microbiological pathogens
- 10. High engraftment for xeno-transplants







Histology of thymus from newborn mice







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NOD
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NOD/SCID

NOG

Basic characteristics ----- Irradiation sensitivity



Survival rates of NOG mice after irradiation

Five mice in each group were irradiated with 2 to 4 Gy with using an X-ray device (MBR-1505R, Hitachi Medical Co., Tokyo) at age of 8 weeks.

Lymphocytes Leakiness in Immunodeficient Mice

Mouse strain	Spleen test	No. of potisive mice/ No. of mice* (range, μg/ml)					
	CD19+	CD3+	CD4+	CD8+	TCRVb	TCRVgd	
C.B-17-scid	6/10	1/10	1/10	0/10	1/10	1/10	
	(0.5 - 6.3)	(9.3)	(6.5)	-	(7.1)	(2.1)	
NOD-scid	0/9	9/9	6/9	8/9	9/9	4/9	
	-	(2.0 - 13.8)	(1.0 - 12.5)	(0.6 - 7.8)	(0.9-12.4)	(0.5 - 1.1)	
NOG	0/18	0/18	0/18	0/18	0/18	0/18	
	-	-	-	-	-	-	
IQI	4/4	4/4	4/4	4/4	4/4	4/4	
	(34.7 - 59.9)	(21.5 - 34.9)	(17.9 - 25.5)	(1.5 - 3.5)	(20.0 - 33.2)	(0.7 - 1.3)	
Mouse strain	Peripheral blood test No. of potisive mice/ No. of mice*						
		range, μg/ml)					
	CD19+	CD3+	CD4+	CD8+	TCRVb	TCRVgd	
C.B-17-scid	4/10	0/10	0/10	0/10	0/10	0/10	
	(0.5 -0. 8)	-	-	-	-	-	
NOD-scid	0/9	2/9	2/9	2/9	4/9	0/9	
		(2.4 - 3.2)	(1.53 - 2.26)	(0.8 - 2.1)	(0.5 - 4.4)		
NOG	0/18	0/18	0/18	0/18	0/18	0/18	
	-	-	-	-	-	-	
IQI	4/4	4/4	4/4	4/4	4/4	3/4	
	(28.1 -45.5)	(14.4 - 31.3)	(10.4 - 23.9)	(2.6 - 5.1)	(13.7 - 30.0)	(0.5 - 1.4)	

* No. of positive mice means the number of mice with more than 0.5% of cells stained, since a value of less than 0.5% was considered as nonspecific staining.

No B cell leakiness in NOG mice



C.B-17-scid: 6-9 months old* C57BL/6 & BALB/c: 12 weeks old

*Ig M+G+A levels in sera of C.B-17- scid were measured in 1989.

Murine cells in bone marrow and spleen



Bone marrow and spleen were obtained from 12 weeks-old NOG mice. Single cell suspensions prepared from them according to the ordinal manner were stained with FITC- or PE-labeled anti-mouse CD45+, CD3+, CD4+, CD8+, CD19+, B220+, CD11b+, CD11c+, Gr-1+ and analyzed with FACSCanto (BD sciences, CA).

Immunological characteristics ----- NK cell defect



NK cell Activity of NOG mice

The NK cell activity was determined by a cytotoxicity test using NK sensitive YAC-1 cells as a target cell. Mice were intraperitoneally inoculated with 100 mg of polyinosinic-polycytidylic acid (poly I:C, SIGMA Chemical Co., St. Louis, MO) to stimulate NK cell activity for 48 hr before assay. Spleen cells were separated from 4 mice of each strain of mice, pooled and co-cultured with ⁵¹Cr-labeld YAC-1 cells as target cells for 4 hr at 37 °C in 5% CO₂. The supernatants harvested were assayed on a gamma counter. The present specific ⁵¹Cr release was calculated using the following formula, where X is the mean experimental release from triplicate wells. Total release (T) was determined from wells with ⁵¹Cr labeled YAC-1 cells and 1H HCl, and spontaneous release (S) was determined from wells with ⁵¹Cr –labeled YAC-1 cells and medium: % specific release = $[(X-S)/(T-S)] \times 100$.

Immunological characteristics ----- Macrophage and Complement activity



Reduced IL-1 production from macrophages

Stimulation of bone-marrow cells

IL-1 production from bone marrow cells

IL-1 production from bone marrow cells stimulated with IFN-g and LPS was determined. Bone marrow cells were cultured with 500 U/ml human rM-CSF, with and without 10 U/ml rat rIFN-g and cultured for 4 days at 37 °C in 5% CO₂. After 4 days, the medium was replaced with fresh medium alone or with medium containing *Escherichia coli* LPS at 10 mg/ml. After an additional 24 hr incubation period, the culture supernatants were harvested and assayed for IL-1a levels using ELISA kits. The amount of IL-1a in the supernatants was expressed as the absorbance at 405 nm.

Defect of hemolytic complement activity in serum



Serum dilution

Complement-dependent hemolytic activity

Complement-dependent hemolytic activity in sera were determined by measurement of ⁵¹Cr released in the supernatants after 30 min incubation of mouse sera and ⁵¹Cr labeled SRBC/anti SRBC antibody conjugates. Spontaneous release (S) was determined from wells with ⁵¹Cr SRBC-Ab conjugate in media. Total release (T) was determined from wells with ⁵¹Cr SRBC-Ab conjugates and 100 ml 2% SDS. Percent specific release = [(X-S)/(T-S)] x 100.

from Ito, M. et al. NOD/SCID/gamma(c)(null) mouse: an excellent recipient mouse model for engraftment of human cells. *Blood* 100, 3175-3182. (2002).



In vitro cytokine production of *L. monocytogenes*-stimulated spleen cells from 3 strains of mice.

Spleen cells were separated after injection of 1 ml of 1 mg/ml Collagenase D solution into the spleen. CD11c⁺ cells were depleted from spleen cells from NOD/Shi-*scid* mice treated with anti-asialo GM1 antiserum, using anti-CD11c antibody labeled magnetic beads, by magnetic cell sorter (MACS). The cell suspension were co-cultured with 10⁷ of heat-killed *L. monocytogenes* for 8 hours at 37 °C. The IFN-g and IL-6 levels in the supernatants were determined using ELISA kits. Asterisk indicates a significant difference (* vs **: P<0.01).

Collaborative studies with Japanese Academic Society



Application

- 1. Infectious disease model
 - HIV-1 infection
 - ATL infection
 - EBV infection model
 - Hodgkin's disease model
- 2. Cancer model
 - Liver metastasis
 - Multiple myeloma
 - Acute myeloid leukemia
- 3. Human tissue or organ model
 - Model with human ovary
 - Model with human liver
 - Model with human endometrium
- 4. Other models
 - GVHD model
 - Efficacy test model for thrombopoietic drugs
 - Safety test for human cell (ES, iPS, genemanipulated cells) transplantation